



OPEN ACCESS

ORIGINAL ARTICLE

Anti-inflammatory effects of PGE₂ in the lung: role of the EP₄ receptor subtype

Mark A Birrell,^{1,2} Sarah A Maher,¹ Bilel Dekkak,¹ Victoria Jones,¹ Sissie Wong,¹ Peter Brook,¹ Maria G Belvisi^{1,2}

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/thoraxjnl-2014-206592>).

¹Faculty of Medicine, Department of Respiratory Pharmacology, National Heart and Lung Institute, Imperial College London, London, UK
²MRC-Asthma UK Centre in Allergic Mechanisms of Asthma

Correspondence to Professor Maria G Belvisi, Faculty of Medicine, Respiratory Pharmacology, National Heart and Lung Institute, Imperial College London, Exhibition Road, London SW7 2AZ, UK; m.belvisi@imperial.ac.uk

Received 20 November 2014
 Revised 16 February 2015
 Accepted 12 March 2015

ABSTRACT

Background Asthma and chronic obstructive pulmonary disease (COPD) are chronic inflammatory diseases of the airway. Current treatment options (long acting β -adrenoceptor agonists and glucocorticosteroids) are not optimal as they are only effective in certain patient groups and safety concerns exist regarding both compound classes. Therefore, novel bronchodilator and anti-inflammatory strategies are being pursued. Prostaglandin E₂ (PGE₂) is an arachidonic acid-derived eicosanoid produced by the lung which acts on four different G-protein coupled receptors (EP₁₋₄) to cause an array of beneficial and deleterious effects. The aim of this study was to identify the EP receptor mediating the anti-inflammatory actions of PGE₂ in the lung using a range of cell-based assays and in vivo models.

Methods and results It was demonstrated in three distinct model systems (innate stimulus, lipopolysaccharide (LPS); allergic response, ovalbumin (OVA); inhaled pollutant, cigarette smoke) that mice missing functional EP₄ (*Ptger4*^{-/-}) receptors had higher levels of airway inflammation, suggesting that endogenous PGE₂ was suppressing inflammation via EP₄ receptor activation. Cell-based assay systems (murine and human monocytes/alveolar macrophages) demonstrated that PGE₂ inhibited cytokine release from LPS-stimulated cells and that this was mimicked by an EP₄ (but not EP₁₋₃) receptor agonist and inhibited by an EP₄ receptor antagonist. The anti-inflammatory effect occurred at the transcriptional level and was via the adenylyl cyclase/cAMP/ cAMP-dependent protein kinase (PKA) axis.

Conclusion This study demonstrates that EP₄ receptor activation is responsible for the anti-inflammatory activity of PGE₂ in a range of disease relevant models and, as such, could represent a novel therapeutic target for chronic airway inflammatory conditions.

INTRODUCTION

Asthma and chronic obstructive pulmonary disease (COPD) are respiratory diseases with an increasing global prevalence that represent a social and economic burden for both industrialised and developing countries. The World Health Organization states that the number of patients with asthma is 300 million and predicts that this figure will rise to 400 million by 2025,¹ while there are 600 million people with COPD worldwide and the disease is predicted to be the third ranked leading cause of death by 2020.¹ Exacerbations are common events in the lives of patients with asthma and COPD. These episodes are often associated with infections

Key messages

What is the key question?

- Prostaglandin E₂ (PGE₂) possesses anti-inflammatory and bronchodilator activity in clinical studies but unfortunately this is accompanied by airway irritancy, so identification of the receptor mediating the beneficial effects could lead to the development of a novel therapeutic for chronic inflammatory diseases characterised by airflow obstruction.

What is the bottom line?

- EP₄ receptor activation is responsible for the anti-inflammatory activity of PGE₂ across a range of inflammatory in vivo models and in human cell-based assays.

Why read on?

- This study identifies for the first time in a range of clinically relevant models the EP₄ receptor and its associated signalling pathway as a target for a dual bronchodilator/anti-inflammatory therapy for the treatment of respiratory diseases such as asthma and chronic obstructive pulmonary disease.

by viruses or bacteria and cause worsening of symptoms, which can be fatal. Often these heightened symptoms are far less responsive to normal treatments and are associated with increased health-care costs and societal impact. There is therefore an urgent need to develop safe and effective therapies for these respiratory diseases.

It has been recognised for many years that prostaglandin E₂ (PGE₂) is produced by many cell types within the lung.² Furthermore, exogenous PGE₂ has been shown to have bronchodilator and anti-inflammatory properties in rodent models³ and human subjects.^{4,5} Thus, PGE₂ could represent an effective treatment, but it is associated with airway irritation and coughing⁶ and has a very short half-life and, as such, has an inappropriate pharmacokinetic profile for general clinical use.

The biological actions of PGE₂ are predominantly mediated by the activation of the four EP receptors, EP₁₋₄.⁷ We and others have recently shown that the bronchodilator properties of PGE₂

To cite: Birrell MA, Maher SA, Dekkak B, et al. *Thorax* Published Online First: [please include Day Month Year] doi:10.1136/thoraxjnl-2014-206592

in human airways are mediated via activation of the EP₄ receptor,⁸ whereas the undesirable triggering of airway sensory nerves appears to be via its activity at the EP₃ receptor.⁹ However, it is still not clear which receptor is involved in mediating the anti-inflammatory effects of PGE₂. Identification of a receptor mediating the anti-inflammatory and bronchodilator effects of PGE₂ without the airway irritant activity would harness the beneficial effects while avoiding the undesirable properties.

To achieve this aim we profiled the inflammatory status of EP receptor knockout (KO) mice in an array of preclinical respiratory disease model systems (endotoxin: innate; allergen: asthma-like; and smoke: COPD-like). To investigate the signalling mechanisms involved, we used in vitro murine and human cell-based assay systems. In vitro assays confirmed the in vivo data which suggested that EP₄ receptor activation is responsible for the anti-inflammatory activity of PGE₂ and that the effect was at the level of cytokine gene transcription via an adenylyl cyclase (AC)/cAMP/cAMP-dependent protein kinase (PKA) axis.

MATERIALS AND METHODS

Role of EP receptors in murine respiratory models

Breeding pairs of C57bl/6 wild type and EP receptor gene (*Ptger1–Ptger4*) KO mice were purchased from Harlan (Bicester, Oxon, UK) and provided by Dr Shuh Narumiya, Kyoto University, respectively. *Ptger1–Ptger3* KO mice were backcrossed on the C57bl/6 at least eight times. *Ptger4*–/– mice do not survive on the C57bl/6 background¹⁰ and were backcrossed on a mixed background of 129/Ola×C57bl/6. Corresponding wild type colonies were generated. Male mice (18–20 g) were generated from established breeding colonies maintained internally. Experiments were performed under Home Office project licence (PPL 70/7212) and procedures adhered to the Animals (Scientific Procedures) Act 1986 and according to the ARRIVE guidelines.¹¹

Mice were challenged with inhaled lipopolysaccharide (LPS) (innate response), ovalbumin (OVA) (allergic: asthma-like) or cigarette smoke as previously described.^{12–14} Briefly, for the innate response model, mice were challenged with an aerosol of endotoxin-free saline (for 30 min) or LPS (1 mg/mL, *Escherichia coli* serotype 0111:B4). For the asthma model, mice were sensitised with OVA (intraperitoneally with alum on days 0 and 14) and on days 24, 25 and 26 mice were intranasally challenged with saline or OVA (50 µg in 50 µL) under isoflurane anaesthesia. For the COPD model, mice were challenged with cigarette smoke or room air for 1 h twice a day for 3 days. Airway cellular inflammation was assessed via differential counting in the lavage and EP mRNA receptor expression determined in the lung tissue (over time at 2, 6, 24 and 48 h after final challenge).

Role of EP receptors in cell-based systems

Cultured murine (J774) and human (THP-1) monocytes (treated with the cyclo-oxygenase inhibitor indomethacin) were challenged with LPS in the presence and absence of PGE₂. Cytokine levels in the culture medium were assessed using ELISA. In parallel experiments, the impact of specific EP receptor agonists EP₁ (ONO-D1-004), EP₂ (ONO-AE1-259), EP₃ (ONO-AE-248), EP₄ (ONO-AE1-329) and antagonists EP₂ (PF-04418948) and EP₄ (ONO-AE3-208) were studied. Concentrations used were selected from previous studies and published literature.^{8–9–15} Dimethyl sulfoxide was used as the vehicle, final concentration 0.001–2% (v/v). EP receptor

expression was determined in THP-1 cells and primary human alveolar macrophages by RT-PCR.

Cytokine production was measured at the mRNA (RT-PCR 2 h post-challenge) and protein (ELISA 22 h post-challenge) level to assess the effect of EP₄ agonist on transcription/translation. To further explore the signalling mechanism, the role of AC (AC activator: forskolin; inactive control: 1,9 dideoxyforskolin) and PKA/exchange proteins activated by cAMP (EPAC) (PKA activator: 6-Bnz-cAMP; EPAC activator: SP-8-pCPT-2'-O-Me-cAMP; and PKA inhibitor: RP-8-CPT-cAMPS) were explored; concentrations were selected from published data and unpublished concentration–response curves.^{16–17} The key experiment was repeated in primary human alveolar macrophages as previously described.¹⁸ Human lung tissue was purchased from the International Institute for the Advancement of Medicine (Edison, New Jersey, USA). In all cases, consent for use of the tissue in scientific research and ethics approval were obtained from the Royal Brompton and Harefield Trust.

Data analysis and statistics

Data are presented as mean±SEM and statistical significance was assumed when $p < 0.05$. N numbers and statistical tests used were the Mann–Whitney test when two datasets were compared and Kruskal–Wallis ANOVA followed by Dunn's multiple comparison for three or more datasets. The specific test used is indicated in the figure legends.

RESULTS

Role of EP receptors in murine respiratory models of airway inflammation

Wild type mice challenged with the various disease relevant stimuli evoked the expected inflammatory responses associated with increases in a particular cell type. In particular, increases in airway neutrophils were observed in the LPS and cigarette smoke models and eosinophils after antigen challenge (figure 1). In the LPS and allergen models, inflammatory cell infiltration was significantly increased in the EP₄ receptor (*Ptger4*–/–) KO mice (with no change in EP_{1–3} KO mice) compared with the wild type control, suggesting a protective role for EP₄ receptor activation (figure 1). To determine if this phenomenon was restricted to these inflammatory stimuli, we compared inflammatory cell infiltration into the airway following cigarette smoke challenge and found that absence of the EP₄ receptor appeared to enhance the inflammatory response (figure 1E).

The EP₄ KO mice, of necessity, were on a different genetic background to the other three KO lines.¹⁰ This difference in genetic background between the studies with EP_{1–3} and EP₄ receptor KO mice could conceivably complicate the data interpretation. Thus, in order to provide evidence that functional changes were associated with the 'disease phenotype' and not specific to the genetic background of the mice, we assessed EP receptor expression in the allergen and LPS models in mice bred on a pure C57bl/6 background. While we could not detect a robust temporal change in the levels of EP_{1–3} receptor mRNA, EP₄ receptor mRNA levels were significantly increased across a number of time points in both inflammatory models (see online supplementary figures S1 and S2).

Anti-inflammatory role of EP receptors in cell-based assay systems

In three different disease relevant in vivo models of airway inflammation we found that the inflammatory response was increased in EP₄ receptor KO mice, suggesting an anti-inflammatory role for endogenous prostaglandins via activation

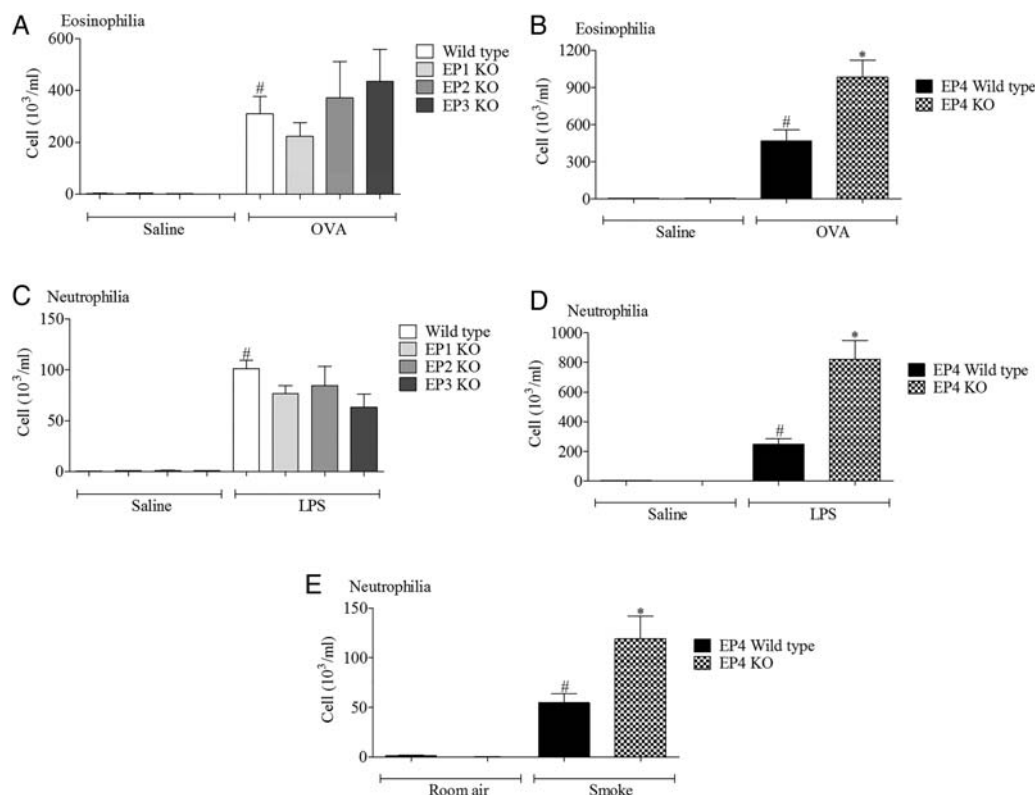


Figure 1 Role of EP receptors in murine respiratory models. (A, B) Eosinophils in lavage fluid from mice sensitised and challenged with ovalbumin (OVA), wild type controls versus EP₁₋₃ knockout (KO) mice (A) or wild type controls versus EP₄ KO mice (B). (C, D) Neutrophils in lavage fluid from mice challenged with lipopolysaccharide (LPS), wild type controls versus EP₁₋₃ KO mice (C) or wild type controls versus EP₄ KO mice (D). (E) Neutrophils in lavage fluid from mice exposed to cigarette smoke, wild type controls versus EP₄ KO mice. Data shown are mean±SEM (n=8). #Statistical significance between control challenge and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between wild type and KO mice (p<0.05, Mann–Whitney test).

of EP₄ receptors. In order to perform more in-depth investigations using pharmacological tools without the appropriate pharmacokinetic profile for in vivo studies and in order to provide

translational data, we performed studies in human cell-based assays. To investigate the anti-inflammatory potential of PGE₂, we examined the effect of exogenously added PGE₂ on

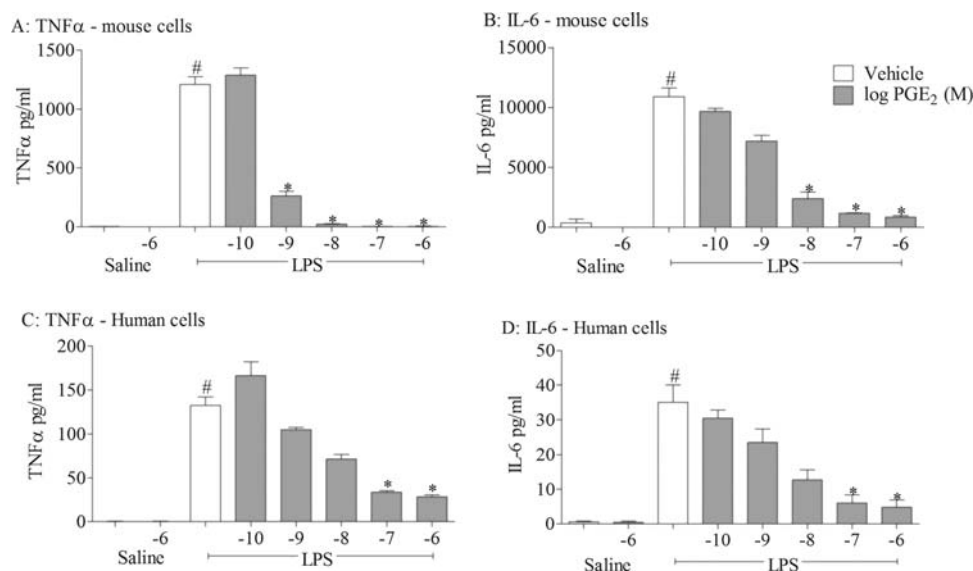


Figure 2 Effect of prostaglandin E₂ (PGE₂) on cytokine release in cell-based assays. Effect of increasing concentrations of PGE₂ on levels of tumour necrosis factor α (TNFα) (A, C) and interleukin 6 (IL-6) (B, D) in culture medium from mouse (J774; A, B) and human (THP-1; C, D) THP-1 cells stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/ml). Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation (p<0.05, Mann–Whitney test). *Statistical significance between stimulation and PGE₂-treated cells (p<0.05, Kruskal–Wallis ANOVA followed by Dunn's multiple comparison test).

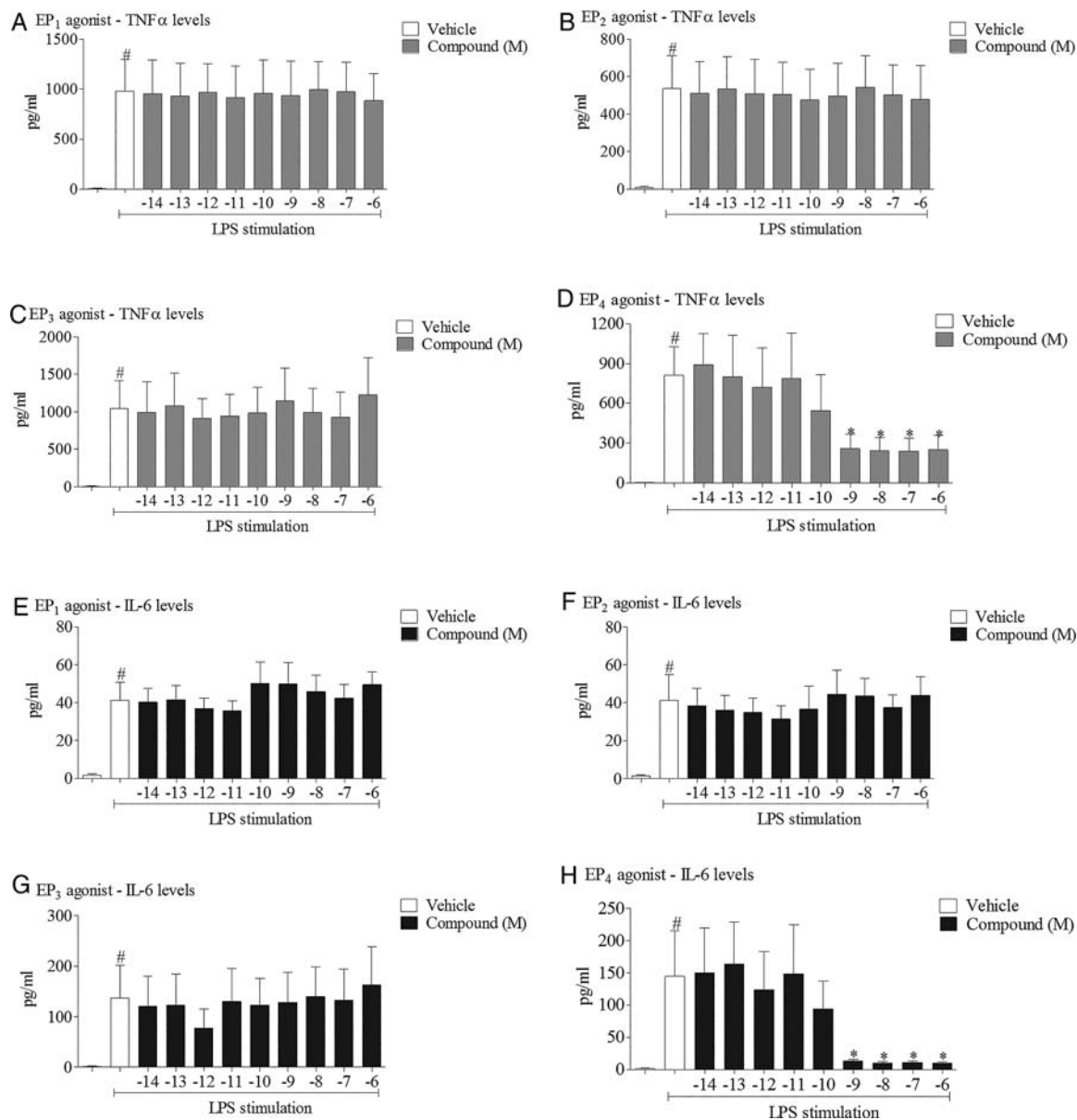


Figure 3 Effect of selective EP receptor agonists on cytokine release. Effect of increasing concentrations of EP receptor agonists (EP₁: ONO-D1-004, EP₂: ONO-AE1-259, EP₃: ONO-AE-248, EP₄: ONO-AE1-329) on levels of tumour necrosis factor α (TNF α) (A–D) and interleukin 6 (IL-6) (E–H) in culture medium from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μ g/ml). Data shown are mean $\pm$ SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation ($p < 0.05$, Mann-Whitney test).

*statistical significance between stimulation and agonist treated cells ($p < 0.05$, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).

inflammatory responses to one of the stimuli used in the murine model, namely LPS. All experiments were performed in the presence of a broad spectrum cyclo-oxygenase inhibitor to block endogenous prostanoid production which may complicate data interpretation. PGE₂ produced a concentration-related inhibition of LPS-induced cytokine production in mouse and human monocytes (figure 2). To investigate which receptor(s) were central to the anti-inflammatory effect seen with PGE₂, we repeated the study using selective EP agonists and found that only the EP₄ receptor agonist (ONO-AE1-329) modulated cytokine levels at concentrations commensurate with the reported nanomolar potency of this ligand at the EP₄ receptor (figure 3D, H). In a parallel experiment, the same EP receptor agonists did not trigger cytokine release under non-stimulated conditions (see online supplementary figure S3). Interestingly, EP₄ receptor mRNA expression was the predominant EP receptor subtype

expressed at the mRNA level in these cells and primary human alveolar macrophages (see online supplementary figure S4). These data suggest that the EP receptor expression profile and the predominance of EP₄ explains why the anti-inflammatory activity of PGE₂ is mediated via the EP₄ receptor.

To confirm that the EP₄ receptor is central to the anti-inflammatory properties of PGE₂, we performed experiments in the presence of selective EP₂ and EP₄ receptor antagonists (PF-04418948 and ONO-AE3-208, respectively) and focused on tumour necrosis factor α (TNF α) production as an example cytokine. The data clearly show that the EP₄ receptor antagonist blocks the anti-inflammatory activity of both the selective EP₄ receptor agonist and PGE₂ itself (figure 4C, D), with no effect of the EP₂ receptor antagonist.

In order to investigate whether the production of TNF α was regulated at the transcriptional or translational level, we had to

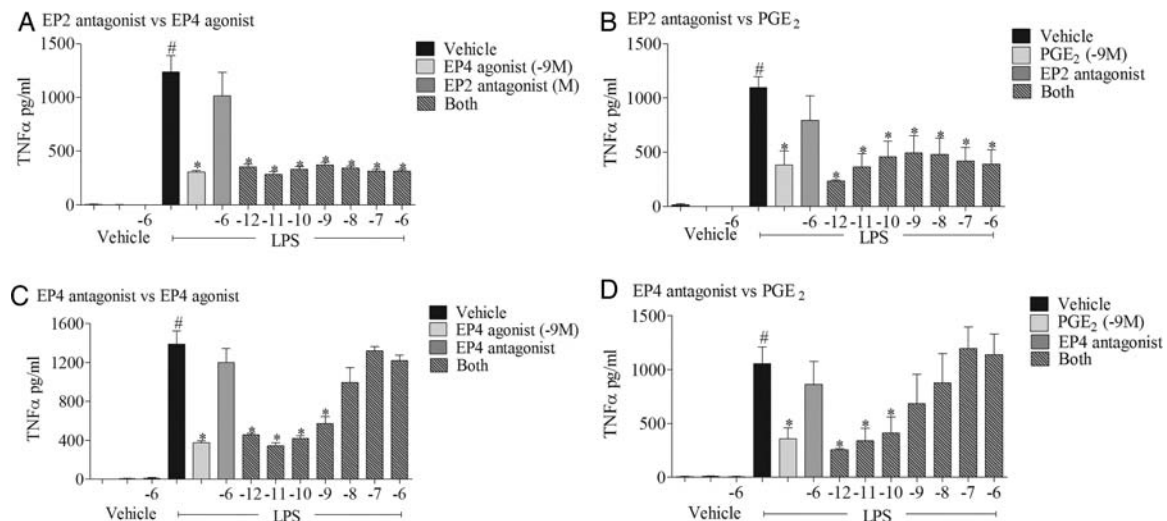


Figure 4 Effect of selective EP receptor antagonists on inhibitory effect of prostaglandin E₂ (PGE₂) and EP₄ agonist-induced cytokine release. Effect of EP₄ receptor agonist (ONO-AE1-329, 1 nM) (A, C) or PGE₂ (1 nM) (B, D) in the presence of increasing concentrations of EP₂ (PF-04418948) or EP₄ (ONO-AE3-208) receptor antagonists on tumour necrosis factor α (TNF α) levels in culture medium from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μ g/mL). Data shown are mean $\pm$ SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation ($p < 0.05$, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells ($p < 0.05$, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).

determine a suitable time point to measure mRNA production. The 2 h time point was selected to measure mRNA and protein production was assessed 22 h after stimulation (figure 5A). The EP₄ receptor agonist modulated both the mRNA and protein levels, which suggests that activation of the receptor modulates cytokine production at the transcriptional level (figure 5C, D).

To explore the signalling mechanisms involved, we investigated the effects of mimicking proposed EP₄ receptor signalling involving the AC-PKA/EPAC pathways. The AC activator, but not its negative control (1,9 dideoxyforskolin, which shares many of the

activities of forskolin but does not activate AC), caused a concentration-related inhibition of cytokine production which was of a similar magnitude to the EP₄ receptor agonist (figure 6A, B). Similarly, an activator of PKA, but not EPAC, mirrored the effect of the EP₄ receptor agonist (figure 6C, D). In order to determine if this proposed pathway was indeed triggered by activating the EP₄ receptor, we employed a selective PKA blocker. The PKA inhibitor caused a concentration-related block of the anti-inflammatory properties of both a PKA activator and the EP₄ receptor agonist (figure 7A, B). Finally, to demonstrate this

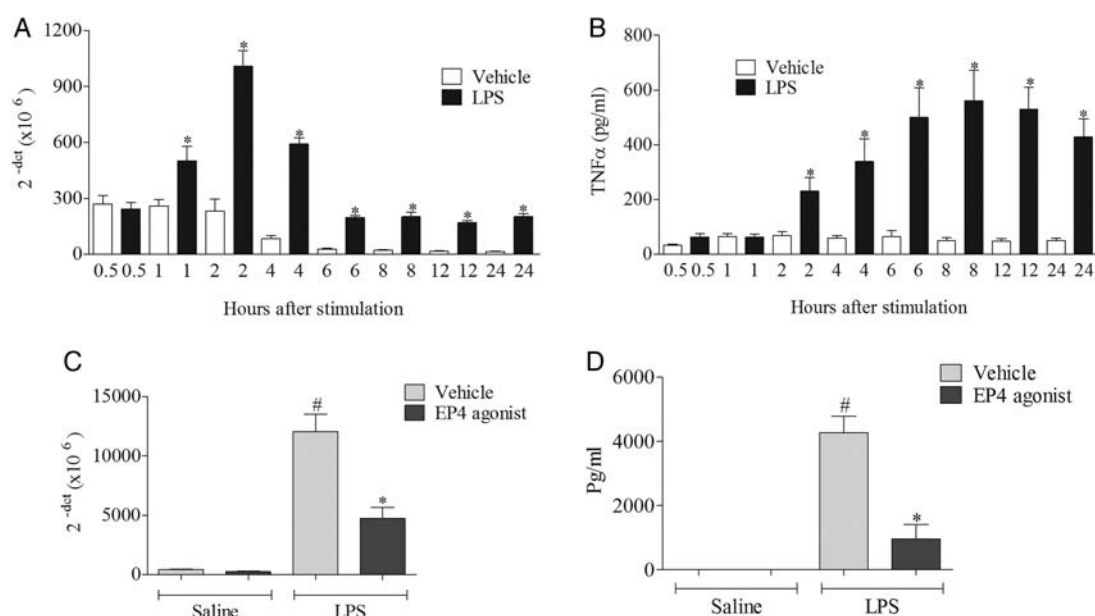


Figure 5 Assessing the effect of EP₄ receptor activation on transcription and translation of tumour necrosis factor α (TNF α). TNF α mRNA (A) and protein production (B) following treatment with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μ g/mL) over time in the culture medium of human (THP-1) monocytes. (C) TNF α mRNA measured 2 h post stimulation and (D) protein measured 22 h post stimulation in the presence of EP₄ receptor agonist (ONO-AE1-329). Data shown are mean $\pm$ SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation ($p < 0.05$, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells ($p < 0.05$, Mann–Whitney test).

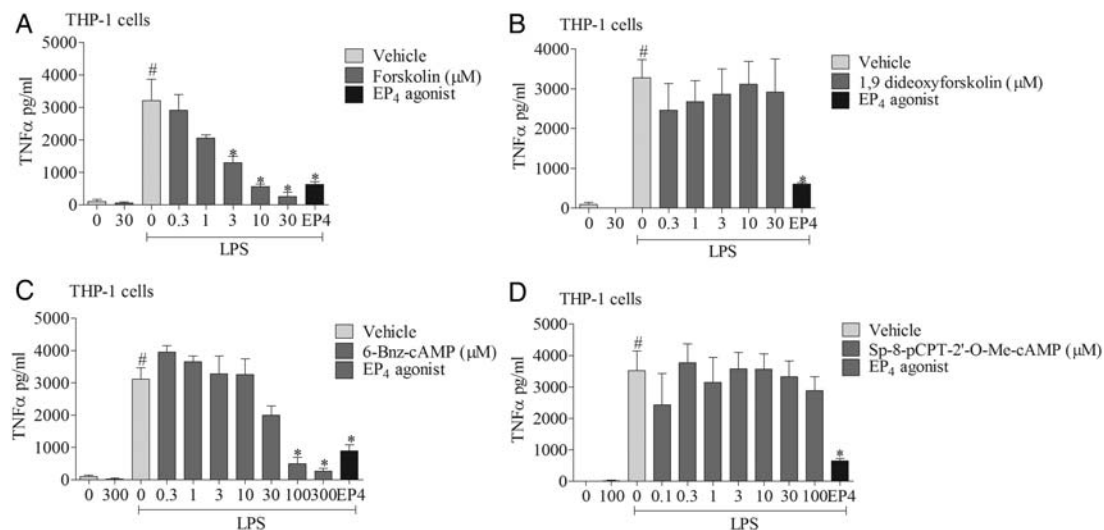


Figure 6 EP₄ receptor activation and downstream signalling pathways. The effect of vehicle, adenylyl cyclase (AC) activator (A), AC activator negative control (B), cAMP-dependent protein kinase (PKA) activator (C), or exchange proteins activated by cAMP (EPAC) activator (D) on tumour necrosis factor α (TNF α) release from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μ g/mL). The EP₄ receptor agonist (ONO-AE1-329, 1 nM) was included as a positive control. Data shown are mean $\pm$ SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation ($p < 0.05$, Mann–Whitney test). *Statistical significance between stimulation and agonist treated cells ($p < 0.05$, Kruskal–Wallis ANOVA followed by Dunn’s multiple comparison test).

pathway was active in primary cells, we repeated the observation in human alveolar macrophages (figure 7C).

DISCUSSION

Current anti-inflammatory therapy for asthma and COPD is focused on the use of glucocorticoids, but their efficacy is limited in COPD and in certain patient subgroups with asthma. Furthermore, there are significant patient compliance issues and safety concerns associated with their long-term use. Existing treatment of both diseases is focused on combination therapies which include a bronchodilator together with the glucocorticoid anti-inflammatory. Therefore, a single treatment with both anti-inflammatory and bronchodilator potential would be a preferred therapeutic option, especially if it could be developed as an oral medication to avoid compliance issues.

The beneficial properties of PGE₂ in the lung have long been recognised, but irritancy issues and the poor pharmacokinetic profile of endogenous bioactive lipid mediators mean that these properties are yet to be fully harnessed. Previous studies have started to define the EP receptors associated with the beneficial and undesirable properties associated with PGE₂ to identify a receptor subtype which could be targeted for the development of a therapeutic. For example, EP₄ receptor activation has been proposed to be involved in PGE₂-induced bronchodilation of human airway smooth muscle and EP₃ receptor activation to trigger airway sensory nerves producing unwanted reflexes such as cough.^{8–9} The aim of this study was to begin to explore which receptor(s) are required for the airway anti-inflammatory properties of PGE₂.

The initial body of work was performed in three *in vivo* murine models with distinct phenotypes (modelling innate, allergic and COPD-like inflammation) and demonstrated that EP₄ receptor KO mice had an enhanced cellular inflammation, suggesting an endogenous anti-inflammatory role for PGE₂ acting on the EP₄ receptor. Supporting these data was the observation that EP₄ receptor gene expression was increased in the lungs after disease relevant challenges in standard (C57bl/6) wild type mice, suggesting that this observation was not specific

to the genetic background of the mice used in the EP₄ comparator studies. Consistent with these findings are the facts that most immune cell types have been reported to express EP₄ receptors,^{7–19–20} and EP₄ receptor activation reduces transendothelial migration of eosinophils and neutrophils^{21–22} and induces suppression in a murine model of aspirin-triggered allergic hyperresponsiveness.²³ In contrast to these observations, a protective role for EP₃ receptor activation has been reported in a murine asthma model.²⁴ Although a beneficial role for the EP₃ receptor is not supported in studies demonstrating that PGE₂—but not the EP₃ receptor agonist sulprostone—exhibits anti-inflammatory effects in an allergic murine model, and in other studies suggesting that EP₃ receptor activation is chemotactic for lung mast cells.²⁵ Alternatively, other studies have reported that EP₂ receptor activation inhibits asthma-related eosinophilic trafficking.^{26–27} Although studies exist supporting a role for alternative receptors in mediating the anti-inflammatory effects of PGE₂, often the study investigators have not had the opportunity to use the wide range of selective pharmacological tools and KO mice that we have been able to employ here. Furthermore, the majority of the published literature regarding the anti-inflammatory role of PGE₂ is in agreement with our data suggesting a role for the EP₄ receptor.

To provide further confirmatory data regarding the role of the EP₄ receptor and to investigate the downstream signalling pathway, we used cell-based systems which are more amenable to study and where data interpretation is not limited by the pharmacokinetic limitations of tool compounds. This approach also enabled translation of findings in murine systems to human cells. Monocyte/macrophage cell types were used as the prevalent cell in the airway and the cell type most likely to have a common role in all three inflammatory responses that were modelled in the mouse. Furthermore, previous studies had demonstrated EP₄ receptor gene expression in murine macrophages,²⁸ the murine monocyte cell line J774 cells²⁹ and human lung macrophages.^{20–30} In these experiments, using one of the stimuli used in the model systems (endotoxin, LPS), PGE₂ also had an anti-inflammatory role demonstrated as the inhibition of

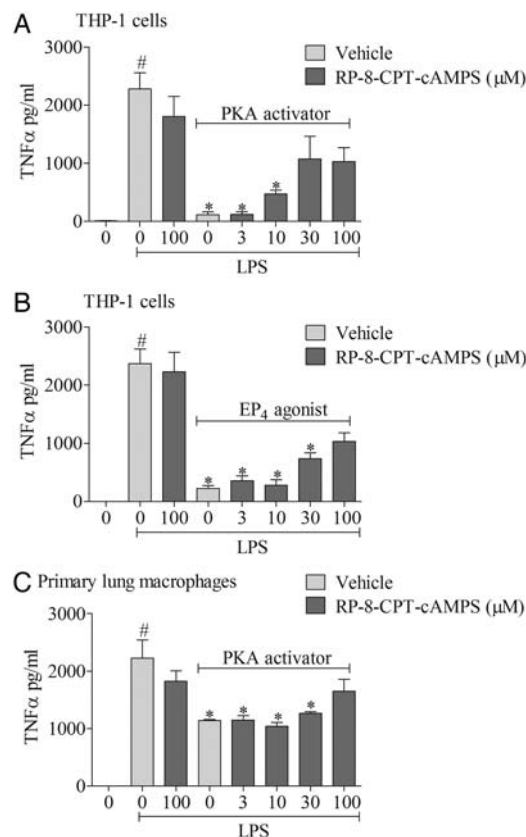


Figure 7 EP₄ receptor activation and downstream signalling pathways. Effect of a cAMP-dependent protein kinase (PKA) activator (6-Bnz-cAMP, 100 μM) plus increasing concentrations of a PKA inhibitor (RP-8-CPT-cAMPS) (A) or EP₄ receptor agonist (ONO-AE1-329, 1 nM) plus the PKA inhibitor (B). PKA activator plus increasing concentrations of PKA inhibitor on tumour necrosis factor α (TNFα) levels in culture medium from human (THP-1) monocytes stimulated with vehicle (saline) or lipopolysaccharide (LPS; 0.1 μg/mL). The effect of a PKA activator plus increasing concentrations of PKA inhibitor (C) in primary human alveolar macrophages stimulated with LPS or vehicle. Data shown are mean±SEM of three experimental runs performed in duplicate. #Statistical significance between vehicle and stimulation ($p<0.05$, Mann-Whitney test). *Statistical significance between stimulation and agonist treated cells ($p<0.05$, Kruskal-Wallis ANOVA followed by Dunn's multiple comparison test).

cytokine release via activation of the EP₄ receptor. These data are consistent with previous studies that have reported a similar role for the EP₄ receptor in a diverse range of cell-based assays.^{19 20 29 31 32} Interestingly, we did not observe an anti-inflammatory role for the EP₂ receptor, as suggested by other studies. In addition, although we failed to note any pro-inflammatory effects of PGE₂, it has been reported to increase levels of inflammatory mediators such as vascular endothelial growth factor through both EP₂ and EP₄ receptors.³³

To investigate how activation of the EP₄ receptor attenuates cytokine production, we probed the reported signalling mechanisms. EP₄ receptors signal via activation of Gαs and AC and a subsequent increase in intracellular cAMP which can then activate PKA or EPAC.⁷ While others have suggested different signalling mechanisms such as EP₄ receptor-associated protein (p105/NF-κB) and Gαi (PI3K/PKC),^{32 34 35} many report that the cAMP/PKA axis is central to the response.⁷ In our cell systems we demonstrated that the anti-inflammatory actions of PGE₂ and the specific EP₄ receptor agonist were mimicked by

AC and PKA activators, but not an EPAC activator, and blocked by a PKA inhibitor. Furthermore, cytokine production was suppressed at the mRNA level, presumably by halting its transcription or by increasing mRNA degradation.

In conclusion, these findings present a convincing dataset that supports the hypothesis that the anti-inflammatory activity of PGE₂ is mediated via activation of the EP₄ receptor and that this mechanism is capable of suppressing airway inflammation that is triggered via an array of stimuli. We have shown that EP₄ receptor KO mice have enhanced inflammation after innate, allergic and COPD-like conditions, and cytokine production from primary human alveolar macrophages is attenuated by an EP₄ agonist in a cAMP/PKA-dependent manner. We suggest that an EP₄ receptor agonist with an appropriate pharmacokinetic profile would harness the beneficial properties of PGE₂ and make an effective anti-inflammatory and bronchodilator for the treatment of respiratory diseases such as asthma and COPD.

Acknowledgements *Ptger1–Ptger4* KO mice and the EP ligands were kindly supplied by Professor Shuh Narumiya, Kyoto University and Professor Takayuki Maruyama, ONO Pharmaceuticals. The EP₂ receptor antagonist was a gift from Dr Nick Pullen, Pfizer.

Contributors Conception and design: MAB and MGB. Data generation, analysis and interpretation: BD, SAM, PB. Drafting the manuscript for important intellectual content: MAB, BD, MGB. All authors reviewed the manuscript and approved the final draft.

Funding SAM and the project consumables were funded by a project grant from the Medical Research Council, (MRC, UK) (SAM, G0800195). VJ was funded by an MRC studentship. PB was supported by a NHLI Trust studentship. EDD was funded by a Wellcome Trust project grant (089301/Z/09/Z) and Boehringer-Ingelheim.

Competing interests None.

Ethics approval Ethical approval was obtained from the Royal Brompton Harefield ethics committee.

Provenance and peer review Not commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt and build upon this work, for commercial use, provided the original work is properly cited. See: <http://creativecommons.org/licenses/by/4.0/>

REFERENCES

- World Health Organization. *Global surveillance, prevention and control of chronic respiratory diseases: a comprehensive approach*. World Health Organization, 2007:12–36.
- Piper P, Vane J. The release of prostaglandins from lung and other tissues. *Ann N Y Acad Sci* 1971;180:363–85.
- Martin JG, Suzuki M, Maghni K, et al. The immunomodulatory actions of prostaglandin E2 on allergic airway responses in the rat. *J Immunol* 2002;169:3963–9.
- Gauvreau GM, Watson RM, O'Byrne PM. Protective effects of inhaled PGE2 on allergen-induced airway responses and airway inflammation. *Am J Respir Crit Care Med* 1999;159:31–6.
- Pavord ID, Wong CS, Williams J, et al. Effect of inhaled prostaglandin E2 on allergen-induced asthma. *Am Rev Respir Dis* 1993;148:87–90.
- Costello JF, Dunlop LS, Gardiner PJ. Characteristics of prostaglandin induced cough in man. *Br J Clin Pharmacol* 1985;20:355–9.
- Woodward D, Jones R, Narumiya S. International Union of Basic and Clinical Pharmacology. LXXXIII: Classification of prostanoid receptors, updating 15 years of progress. *Pharmacol Rev* 2011;63:471–538.
- Buckley J, Birrell MA, Maher SA, et al. EP4 receptor as a new target for bronchodilator therapy. *Thorax* 2011;66:1029–35.
- Maher SA, Birrell MA, Belvisi MG. Prostaglandin E2 mediates cough via the EP3 receptor: implications for future disease therapy. *Am J Respir Crit Care Med* 2009;180:923–8.
- Segi E, Sugimoto Y, Yamasaki A, et al. Patent ductus arteriosus and neonatal death in prostaglandin receptor EP4-deficient mice. *Biochem Biophys Res Commun* 1998;246:7–12.
- Kilkenny C, Browne W, Cuthill I. The ARRIVE guidelines. *RequartoCom* 2010;8:1–2.
- Eltom S, Stevenson CS, Rastrick J, et al. P2X7 receptor and caspase 1 activation are central to airway inflammation observed after exposure to tobacco smoke. *PLoS One* 2011;6:e24097–e24097.

Respiratory research

- 13 Rastrick JMD, Stevenson CS, Eltom S, *et al.* Correction: cigarette smoke induced airway inflammation is independent of NF- κ B signalling. *PLoS ONE* 2013;8:e54128.
- 14 Birrell MA, De Alba J, Catley MC, *et al.* Liver X receptor agonists increase airway reactivity in a model of asthma via increasing airway smooth muscle growth. *J Immunol* 2008;181:4265–71.
- 15 Birrell MA, Maher SA, Buckley J, *et al.* Selectivity profiling of the novel EP2 receptor antagonist, PF-04418948, in functional bioassay systems: atypical affinity at the guinea pig EP2 receptor. *Br J Pharmacol* 2013;168:129–38.
- 16 Roscioni SS, Dekkers BGJ, Prins AG, *et al.* cAMP inhibits modulation of airway smooth muscle phenotype via the exchange protein activated by cAMP (EPAC) and protein kinase A. *Br J Pharmacol* 2011;162:193–209.
- 17 Gjertsen BT, Mellgren G, Otten A, *et al.* Novel (Rp)-cAMPS analogs as tools for inhibition of cAMP-kinase in cell culture. Basal cAMP-kinase activity modulates interleukin-1 β action. *J Biol Chem* 1995;270:20599–607.
- 18 Birrell MA, Wong S, Catley MC, *et al.* Impact of tobacco-smoke on key signaling pathways in the innate immune response in lung macrophages. *J Cell Physiol* 2008;214:27–37.
- 19 Nataraj C, Thomas DW, Tilley SL, *et al.* Receptors for prostaglandin E(2) that regulate cellular immune responses in the mouse. *J Clin Invest* 2001;108:1229–35.
- 20 Ratcliffe MJ, Walding A, Shelton PA, *et al.* Activation of E-prostanoid4 and E-prostanoid2 receptors inhibits TNF-alpha release from human alveolar macrophages. *Eur Respir J* 2007;29:986–94.
- 21 Konya V, Philipose S, Bálint Z, *et al.* Interaction of eosinophils with endothelial cells is modulated by prostaglandin EP4 receptors. *Eur J Immunol* 2011;41:2379–89.
- 22 Konya V, Üllen A, Kampitsch N, *et al.* Endothelial E-type prostanoid 4 receptors promote barrier function and inhibit neutrophil trafficking. *J Allergy Clin Immunol* 2013;131:532–40.
- 23 Shi M, Shi G, Tang J, *et al.* Myeloid-derived suppressor cell function is diminished in aspirin-triggered allergic airway hyperresponsiveness in mice. *J Allergy Clin Immunol* 2014;134:1163–74.
- 24 Kunikata T, Yamane H, Segi E, *et al.* Suppression of allergic inflammation by the prostaglandin E receptor subtype EP3. *Nat Immunol* 2005;6:524–31.
- 25 Weller CL, Collington SJ, Hartnell A, *et al.* Chemotactic action of prostaglandin E2 on mouse mast cells acting via the PGE2 receptor 3. *Proc Natl Acad Sci USA* 2007;104:11712–17.
- 26 Sturm EM, Schratl P, Schuligoi R, *et al.* Prostaglandin E2 inhibits eosinophil trafficking through E-prostanoid 2 receptors. *J Immunol* 2008;181:7273–83.
- 27 Zasłona Z, Okunishi K, Bourdonnay E, *et al.* Prostaglandin E2 suppresses allergic sensitization and lung inflammation by targeting the E prostanoid 2 receptor on T cells. *J Allergy Clin Immunol* 2014;133:379–87.
- 28 Pavlovic S, Du B, Sakamoto K, *et al.* Targeting prostaglandin E2 receptors as an alternative strategy to block cyclooxygenase-2-dependent extracellular matrix-induced matrix metalloproteinase-9 expression by macrophages. *J Biol Chem* 2006;281:3321–8.
- 29 Katsuyama M, Ikegami R, Karahashi H, *et al.* Characterization of the LPS-stimulated expression of EP2 and EP4 prostaglandin E receptors in mouse macrophage-like cell line, J774.1. *Biochem Biophys Res Commun* 1998;251:727–31.
- 30 Kubo S, Takahashi HK, Takei M, *et al.* E-prostanoid (EP)2/EP4 receptor-dependent maturation of human monocyte-derived dendritic cells and induction of helper T2 polarization. *J Pharmacol Exp Ther* 2004;309:1213–20.
- 31 Zhong WW, Burke PA, Drotar ME, *et al.* Effects of prostaglandin E2, cholera toxin and 8-bromo-cyclic AMP on lipopolysaccharide-induced gene expression of cytokines in human macrophages. *Immunology* 1995;84:446–52.
- 32 Minami M, Shimizu K, Okamoto Y, *et al.* Prostaglandin E receptor type 4-associated protein interacts directly with NF-kappaB1 and attenuates macrophage activation. *J Biol Chem* 2008;283:9692–703.
- 33 Bradbury D, Clarke D, Seedhouse C, *et al.* Vascular endothelial growth factor induction by prostaglandin E2 in human airway smooth muscle cells is mediated by E prostanoid EP2/EP4 receptors and SP-1 transcription factor binding sites. *J Biol Chem* 2005;280:29993–30000.
- 34 Wall EA, Zavzavadjian JR, Chang MS, *et al.* Suppression of LPS-induced TNF-alpha production in macrophages by cAMP is mediated by PKA-AKAP95-p105. *Sci Signal* 2009;2:ra28.
- 35 Luschni-Schratl P, Sturm EM, Konya V, *et al.* EP4 receptor stimulation down-regulates human eosinophil function. *Cell Mol Life Sci* 2011;68:3573–87.

THORAX

Anti-inflammatory effects of PGE₂ in the lung: role of the EP₄ receptor subtype

Mark A Birrell, Sarah A Maher, Bilel Dekkak, Victoria Jones, Sissie Wong, Peter Brook and Maria G Belvisi

Thorax published online May 4, 2015

Updated information and services can be found at:

<http://thorax.bmj.com/content/early/2015/05/03/thoraxjnl-2014-206592>

These include:

Supplementary Material

Supplementary material can be found at:

<http://thorax.bmj.com/content/suppl/2015/03/31/thoraxjnl-2014-206592.DC1.html>

References

This article cites 34 articles, 13 of which you can access for free at:

<http://thorax.bmj.com/content/early/2015/05/03/thoraxjnl-2014-206592#BIBL>

Open Access

This is an Open Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt and build upon this work, for commercial use, provided the original work is properly cited. See:

<http://creativecommons.org/licenses/by/4.0/>

Email alerting service

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections

Articles on similar topics can be found in the following collections

[Open access](#) (140)
[Inflammation](#) (937)
[Environmental issues](#) (219)
[Asthma](#) (1639)
[Health education](#) (1084)
[Smoking](#) (923)
[Tobacco use](#) (924)

Notes

To request permissions go to:

<http://group.bmj.com/group/rights-licensing/permissions>

To order reprints go to:

<http://journals.bmj.com/cgi/reprintform>

To subscribe to BMJ go to:

<http://group.bmj.com/subscribe/>